## Review

# Understanding gastrointestinal perfusion in critical care: so near, and yet so far

Gareth Ackland, Michael PW Grocott and Michael G Mythen

Centre for Anaesthesia, University College London, London, UK

Received: 1 August 2000 Accepted: 8 August 2000 Published: 1 September 2000 Crit Care 2000. 4:269-281

© Current Science Ltd (Print ISSN 1364-8535; Online ISSN 1466-609X)

#### **Abstract**

An association between abnormal gastrointestinal perfusion and critical illness has been suggested for a number of years. Much of the data to support this idea comes from studies using gastric tonometry. Although an attractive technology, the interpretation of tonometry data is complex. Furthermore, current understanding of the physiology of gastrointestinal perfusion in health and disease is incomplete. This review considers critically the striking clinical data and basic physiological investigations that support a key role for gastrointestinal hypoperfusion in initiating and/or perpetuating critical disease.

Keywords: gastric tonometry, sepsis, shock, splanchnic circulation

### Introduction

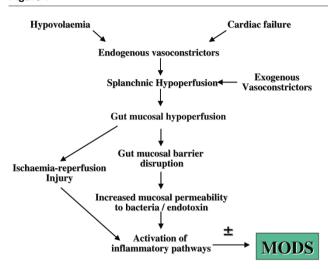
Largely circumstantial evidence continues to implicate the gastrointestinal tract in the pathogenesis of the systemic inflammatory response syndrome (SIRS) and multiple organ dysfunction syndrome. The present review explores why the role of gastrointestinal perfusion has become an important focus of critical care and anaesthesiology research. Specifically, we consider conflicting, unresolved clinical data put forward in support of the idea that the gut is the 'motor of critical disease'. We also consider why current, albeit incomplete, understanding of gastrointestinal circulatory physiology supports this concept.

The idea that the gastrointestinal tract provides the 'spark' and/or 'fuel' for critical disease has been pursued since the series of clinical studies conducted in the 1960s by

Fine and coworkers [1,2], who proposed that a gut-mediated factor, perhaps endotoxin, contributed to sepsis. However, although this attractive idea has gained further support [3,4], pinpointing the exact role of the gastrointestinal tract has proved complex.

One model proposed to explain the involvement of the gut in this process is a two-step mechanism (Fig. 1). First, gastrointestinal perfusion and therefore tissue oxygenation is compromised. Then, as a result of tissue damage, disruption of the mucosal barrier and access to the systemic circulation of toxic entities occurs. The entities proposed include bacteria, bacterial components (eg endotoxin) and chemicals normally found in the bowel lumen. An alternative concept is that the second step involves ischaemia—reperfusion injury of a large viscus, the gastrointestinal

Figure 1



The 'gut hypothesis' for the pathogenesis of critical illness. MODS, multiple organ dysfunction syndrome.

tract, with consequent massive release of cytokines and other proinflammatory mediators. The present review does not focus on the controversies that surround translocation or increased permeability of the gastrointestinal mucosa, or on the evidence for a mechanism that involves ischaemia—reperfusion.

In the clinical setting the results of a number of studies are offered as an example of evidence for the 'gut hypothesis' (Fig. 1). The majority of these studies used gastric tonometry - the only clinical tool for monitoring gastrointestinal perfusion that is widely available at present. Gastric tonometry, using gastric intramucosal pH (pHi) as an index of gastric perfusion, is a highly sensitive but relatively nonspecific predictor of outcome after high-risk major surgery [5], cardiac surgery [6], in a cross-section of patients admitted to the intensive care unit (ICU) [7-9] and in ICU patients with sepsis [10] or acute circulatory failure [11]. However, these data do not establish a causal role for gut hypoperfusion in these situations. The link between gastric perfusion and abnormal tonometry-derived variables is complex. If one accepts that tonometry data reflect perfusion state, abnormal perfusion may still represent an epiphenomenon rather than a causative mechanism. In many ways these clinical studies in the critically ill and high-risk surgical patient serve only to highlight our incomplete physiological understanding of the gastrointestinal tract. We review persuasive data from both human-based and laboratory-based studies that suggest that the splanchnic circulation is important in both health and disease, maintaining regulatory mechanisms that are not obviously linked to gastrointestinal homeostasis alone. Before presenting this data, we briefly review the laboratory and clinical techniques that have been used for the

assessment of gastrointestinal perfusion. A particular focus is on gastric tonometry, the only technology that has accumulated a substantial body of clinical data.

### Assessment of gastrointestinal perfusion

A number of techniques are available for the assessment of gastrointestinal perfusion. Several methods measure portal blood flow or total liver blood flow either directly or indirectly. These include plasma indocyanine green clearance [12] and portal vein catheterization with measurement of blood flow blood flow, oxygen saturation and lactate [13]. Although these techniques have contributed to our understanding of basic physiology, they are not widely used clinically and, in addition, measure total hepatosplanchnic perfusion. We do not focus on these methods any further.

A number of techniques that have not reached the clinical arena are utilized widely in research in this field, and are mentioned in this context throughout the present review. These include Doppler flowmetry of both individual mesenteric vessels and of the serosa and mucosa of the gut [14], reflectance spectrophotometry to index gut mucosal haemaglobin concentration and saturation [15], and the use of oxygen electrodes to assess tissue oxygen levels in the colon [16]. A gut oximeter attached to the antimesenteric boder of the intestine has also been used in animal studies [17]. Radioactive, colour-labelled or fluorescent microspheres can be used in animal studies; when the animal is killed at the end of the study, the distribution of the spheres quantifies relative blood flow to different tissue beds against a reference level [18].

The only practical technique for assessing gastrointestinal perfusion that has entered clinical practise is gastrointestinal tonometry for the measurement of gut intraluminal  $CO_2$  (Fig. 2). It is worth at this point exploring the relationship between gastrointestinal intraluminal  $CO_2$  and blood flow.

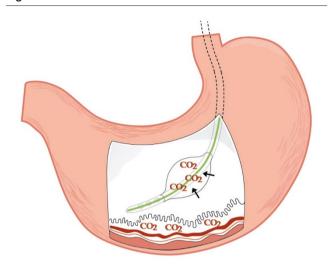
The assumption that intraluminal gut CO2 is elevated when local perfusion is compromised is based on the concept that in situations where gastrointestinal perfusion is reduced oxygen delivery falls below a critical level, resulting in anaerobic cellular metabolism that leads to local lactic acidosis and generation of CO2. An alternative or additional explanation may be inadequate washout of CO<sub>2</sub> due to low flow. When gastrointestinal blood flow is reduced by restriction of superior mesenteric artery (SMA) blood flow in the absence of the hormonal milieu that occurs with systemic hypovolaemia, mucosal pH decreases (CO2 increases) only when flow is less than 50% of baseline [19]. However, this relationship may not hold in hypovolaemia and shock, where vasoactive mediators released in response to decreased intravascular volume are likely to have significant effects on the microcirculation.

Temperature can be an additional confounding factor when the CO2 is measured in the gaseous phase in the stomach, and this is indexed against arterial CO2 measured in the liquid phase. If the two samples are at different temperatures, a methodological error is introduced [20]. It has been suggested that in some cases the Haldane effect may be responsible for increased CO<sub>2</sub> levels in situations of increased oxygen extraction in the absence of decreased perfusion [21]. Clearly local metabolic factors that alter the position of the haemaglobin CO<sub>2</sub> dissociation curve could result in changes in measured gastric CO<sub>2</sub> in the absence of any alteration in local CO<sub>2</sub> production. Although the assumption is made that the CO<sub>2</sub> is of mucosal origin, and this is supported by histological damage to the mucosa in shocked patients, it is possible that the CO2 could be derived from the serosal or muscular levels of the gastrointestinal tract. There are some data to suggest that altered substrate metabolism in the gastrointestinal mucosa may also influence CO<sub>2</sub> production. A lower gastric pHi was observed in swine that were haemorrhaged and resuscitated with a haemaglobin substitute presented in a maltose-containing preparation than those that were resuscitated with a nonsugar-containing preparation [22]. However, hydoxyethyl starch presented in a glucose-containing carrier solution produced less derangement of pHi than the same starch presented in a saline carrier (Wilkes NJ, Woolf R, Mutch M, Stephens R, Mooney L, Mallett SV, Peachey T, Mythen MG, unpublished data).

The development of gastric tonometry as a practical clinical technique has been limited by both methodological drawbacks and problems of interpretation. The original technique of manual saline tonometry was limited by the inconvenience of having to obtain and process samples manually, by slow equilibration times and by errors associated with measurement in the blood gas analyzer. The newer technique of automated semicontinuous air tonometry uses infrared spectrophotometry to measure CO<sub>2</sub>. This system also has the advantage that the partial carbon dioxide tension (Pco<sub>2</sub>) in the intragastric balloon equilibrates more rapidly with the Pco<sub>2</sub> in the stomach, and accurate readings are available within 30 min of commencing monitoring.

The vast majority of clinical outcome and intervention studies use the derived index of gastric pHi. This is obtained by using a formula to produce a value that is claimed to be representative of the tissue pH in the gastric mucosa. However, this assumes that the tissue bicarbonate is equivalent to the arterial bicarbonate. If this is not so, then the derived pH will be inaccurate. Another way of considering this is that the tissue CO<sub>2</sub> signal is being confounded by systemic acid-base disturbances (eg metabolic acidosis) that are known to be independently predictive of outcome.

Figure 2



To nometry in the stomach.  $\ensuremath{\mathrm{CO}}_2$  diffuses into the gastric to nometer balloon.

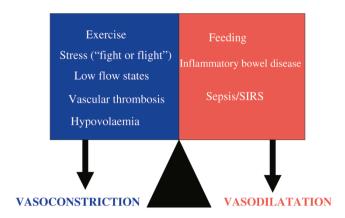
Recent consensus is that presenting the result as the arithmetic difference between the  $PCO_2$  measured in the stomach and the arterial or end-tidal  $PCO_2$ , the  $CO_2$  gap (gastric  $PCO_2$  – arterial  $PCO_2$ , or gastric  $PCO_2$  – end-tidal  $PCO_2$ ), will avoid the problems outlined above [23]. Confirmation that the established clinical correlates with pHi are also true for the  $CO_2$  gap is awaited.

Assessment of gut luminal  $\mathrm{CO}_2$  can be achieved using other techniques and at other sites. Tonometry has also been conducted in the colon [24] in human clinical studies and in the oesophagus [25] in animals in an attempt to develop an easily accessible site for assessing gastro-intestinal perfusion. The sublingual mucosa is an attractive site for clinical measurement because of its ease of access when compared with other parts of the gastro-intestinal tract. However the anatomical basis of sublingual blood flow is significantly different from that of the more distal gut, and it is unclear whether this area has the same susceptibility to hypoperfusion as other gut regions in times of stress. Sublingual capnometry using a  $\mathrm{CO}_2$  electrode placed on the sublingual mucosa has been investigated in humans with limited success [26].

# An overview of the physiology of gastrointestinal perfusion

Under normal circumstances, in addition to the fundamental role of the splanchnic circulation in maintaining liver and gut perfusion to maintain mucosal integrity, the splanchnic bed also acts as a 'circulatory sink' [27]. The redistribution of blood flow that occurs during feeding and exercise are routine haemodynamic challenges for the splanchnic circulation. Exploration of how splanchnic perfusion copes at

Figure 3



A dynamic balance between vasodilatation and vasoconstriction in the gastrointestinal blood supply exists during both health and disease.

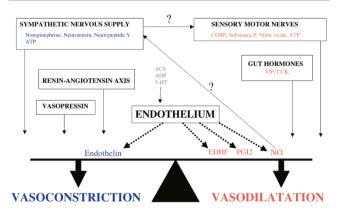
these extremes of normal homeostatic function illustrates the regulatory mechanisms at play (Fig. 3).

We concentrate on splanchnic rather than hepatic perfusion [28], the microvascular perfusion of which has been considered in detail elsewhere [29]. Unless stated otherwise, the studies quoted were conducted using laboratory animal models.

The hepatosplanchnic circulation receives 30% of total cardiac output. With increasing age, splanchnic blood flow declines both absolutely and as a fraction of total cardiac output [30]. Splanchnic anatomy is described in detail elsewhere [31]. Briefly, the mesenteric circulation consists of the muscularis propria, submucosa and mucosa, which are arranged in parallel [32]. Resistance arterioles regulate blood flow to the splanchnic bed, so at constant hydrostatic pressure flow is inversely proportional to resistance. Although these arterioles partake in a markedly less impressive autoregulatory system than in the kidney or brain, they do enable a partial compensation for falls in blood flow [33]. The tone of these vessels depends on the complex balance between neurally mediated sympathetic vasoconstriction, the local action of vasoregulatory substances that are under the influence of the apparently paradoxically named 'sensory-motor' nerves, the parasympathetic cholinergic nerve supply, the enteric nervous system and endothelialderived agents [34] (Fig. 4).

In most models, norepinephrine (noradrenalin) is the key sympathetic-mediated vasoconstrictor acting with the cotransmitters ATP [35] and neuropeptide Y [36], the latter contributing to perhaps 30% of sympathetic vasoconstriction [37]. The vasodilatory calcitonin gene-related

Figure 4



A complex interplay of neural, hormonal and endothelial-derived factors regulates the balance of gastrointestinal perfusion between vasodilatation and vasoconstriction. Question marks indicate possible interactions; dashed lines indicate endothelium-derived production. ACh, acetylcholine; CCK, cholecystokinin; CGRP, calcitonin generelated peptide; EDHF, endothelium-derived hyperpolarizing factor; 5-HT, 5-hydroxytryptamine; PG, prostaglandin; VIP, vasoactive intestinal peptide.

peptide [38] is the main neurotransmitter released at sensory-motor nerves, among many other putative agents. The enteric nervous system includes the nonadrenergic noncholinergic system that supplies perivascular myenteric nerves [39]; nitric oxide (NO) is a putative neurothis system, in addition to transmitter in well-established endothelium-derived role in maintaining basal vascular tone [40]. NO inhibits the synthesis [41] and potent vasoconstrictor action [42] of another endothelial-derived factor, endothelin-1 [43], which belongs to a family of cytokines that exhibit many other roles [44]. Infusion of endothelin-1 produces mesenteric vasoconstriction in the rat, an effect that is attenuated by bosentan, an endothelin-receptor antagonist [45]. Inhibition of NO synthesis not only reveals endothelin to have a tonic pressor role [46], but also increases intestinal epithelial permeability [47]. Human studies show that NG-monomethyl-L-arginine endothelin-1 [48] and (L-NMMA) both reduce splanchnic blood flow, but prior administration of L-NMMA prevents the vasoconstriction normally seen with endothelin-1 [40]. In damaged or absent endothelium in which there is impaired NO production, sympathetic-mediated vasoconstriction has been reported to be augmented, whereas paradoxical vasoconstriction is seen on application of established vasodilatory agents [49]. Such pharmacological interplay may be part of an even more complex system, given that there is evidence for neuromodulation occurring between NO, sympathetic other sensory-motor neurotransmitters and [38,50,51]. Furthermore, there is intriguing data indicating that gastric perfusion can be altered by flow characteristics,

and not simply by volume status. Pulsatile cardiopulmonary bypass results in reduced disturbance in pHi as compared with nonpulsatile cardiopulmonary bypass [52].

Recent biomechanical modelling, based on morphometric mapping of the mesentery, indicated that approximately 40% of the mesenteric circulation is contained in venules, which represent the bulk of the mesenteric microcirculation [53]. Precapillary and postcapillary sphincters determine the tone of these capacitance vessels [54]. The combined action of such capillary sphincters and the resistance arterioles effects intraorgan redistribution [55].

Splanchnic oxygen consumption is 20-35% of total body oxygen consumption [56]. In general, animal models show that oxygen consumption is maintained, even at substantially lower blood flow, by the ability to increase oxygen extraction; only at very low blood flow is oxygen uptake dependent on blood flow [57,58]. This reserve is facilitated by microvascular adaptation; a relatively underperfused, extensive network of collateral capillary beds [59] becomes an additional conduit during periods of decreased oxygen delivery [60]. Mucosal permeability may therefore be protected to a large degree, only becoming compromised when oxygen uptake is below 50% of control [61]. More recent data from human studies [62] suggest that oxygen supply dependency may occur with as little as 30% reduction in gastrointestinal blood flow, with mucosal supply dependency (identified using continuous flow gastric tonometry) occurring before global splanchnic supply dependency can be identified (using portal venous CO2 measurement). Data from studies in humans using tonometry suggest that the mucosa may respond differently to alternative causes of reduction in oxygen delivery. Although stagnant hypoxia is readily detected [63], sensitivity to anaemic hypoxia seems to be much lower [64].

#### Feeding

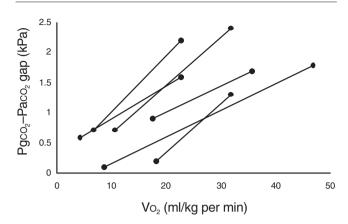
The anticipation and ingestion of food results in neurally mediated sympathetic increases in heart rate, cardiac output, plasma norepinephrine levels and peripheral (forearm) vascular resistance [65]. Within 15 min of food ingestion, SMA blood flow can double from 500 to 1000 ml/min [66], depending on caloric load [67], food volume and type. For example, oral alcohol causes an increased SMA blood flow compared with alcohol-free control [68]. Elevation in SMA blood flow correlates closely with amino-terminal neurotensin and norepiniphrine [67]. Sensory-efferent/motor nerves may play a key role in 'fine-tuning' this process; distension of the large intestine induces SMA smooth muscle hyperpolarization [69] and hence increased blood flow. This may partly explain the observation that the site of feeding can produce differential mesenteric blood flow effects [70]. Fasting for 1 day produces significant mucosal atrophy in the rat [71].

Despite the gut being the largest endocrine organ in the body, the local and cardiovascular effects of many of the gut-derived hormones that are stimulated by feeding have been described relatively recently, and their roles remain unclear [72]. Established humoral agents including vasoactive intestinal peptide [73] and cholecystokinin [74] promote vasodilatation. During ingestion, systemic blood pressure and cardiac output are maintained by increased sympathetic drive, a feature that is absent in patients with autonomic failure [66]. Within 5-30 min after a meal, all cardiovascular responses to feeding subside, except that increased mesenteric blood flow is sustained whereas skeletal muscle blood flow decreases in resting animals [75]. Thus, digestion is accompanied by an increase in total body, myocardial, splanchnic and intestinal oxygen consumption.

The increased metabolic demands of active absorption are proportionally greater than the increases in splanchnic blood flow, suggesting that the active gut may incur an oxygen debt in the same way that we consider the whole body does during exercise and physiological stress. Although the exact mechanism of the absorptive small intestine villi is disputed across species [76-78], numerous authors support the idea that these villi are particularly susceptible to deleterious circulatory or hypoxic changes [79]. The villi exhibit an oxygen countercurrent exchange mechanism, producing relative hypoxia at the luminal tip compared with at its base, even under normal conditions [80,81]. The potential vulnerability of such relatively hypoxic tissue may be potentiated by the villus architecture, which theoretically promotes the phenomenon of plasma skimming, and hence lower haematocrit and oxygen delivery [82], although experimentally this is not seen at reduced perfusion pressures [83].

In the clinical setting there is evidence that the increased metabolic requirements of an absorbing gut coupled with inadequate perfusion due to hypovolaemia or vascular insufficiency may result in a critical imbalance between oxygen supply and demand. The clinical syndrome of mesenteric angina occurring after oral intake is well recognized. Angiography of some of these patients reveals critical arterial stenosis. In patients with stenoses a proportion have an increased difference between their arterial and intragastric Pco2 (an elevated Pco2 gap) at rest, suggesting poor perfusion. Tonometry during moderate exercise demonstrated an abnormal  ${\rm CO_2}$  gap in all patients with stenoses in one study, and this abnormality was absent in control individuals [84]. Revascularization resulted in normalization of perfusion in all patients. The clinical importance of this in a critical care setting is highlighted by case reports that describe patients who have sustained massive bowel infarction soon after initiation of enteral feeding.

Figure 5



Gastric-arterial CO<sub>2</sub> (Pgco<sub>2</sub>-Paco<sub>2</sub>) gap before and after exercise to high oxygen consumption (Vo<sub>2</sub>). Data from Chieverley-Williams S, Hurley R, Cox M, McCorkell S, Grocott MPW, Goldstone J and Mythen MG (unpublished data).

Despite the current consensus that early enteral feeding is beneficial [85,86], the clinical evidence is largely drawn from subset analyses of randomized controlled trials [87], which did not demonstrate an overall outcome benefit, but did show a reduction in specific complications. The possibility that unidentified complications are increased in other subsets cannot be discounted in view of the equivocal overall outcome. There may exist a subset of patients in whom the increased metabolic demands incurred by active absorption of nutrients and increased motility result in critical ischaemia and infarction. Case reports of smallbowel necrosis after jejunal feeding offer support for this idea [88]. It is interesting to speculate whether this group could be identified by tonometry, perhaps in combination with a test of gastric functional response. Good experimental support for this idea is provided in a canine model in which the entire vascular supply to the jejunoileum was isolated [19]. Decreased SMA flow during nutrient delivery only to the jejunum resulted in increased mucosal perfusion at that level (as measured by laser Doppler flowmetry and reflectance spectrophotometry). However, concomitantly, perfusion to the distal ileum was reduced. Thus, a redistribution of mesenteric blood occurred, or 'intramesenteric steal' as coined by the authors.

#### **Exercise**

In contrast to feeding, acute exercise is associated with large increases in cardiac and active skeletal muscle blood flows, but reduced blood flow to skin, kidneys and organs perfused by the splanchnic circulation [89]. Using SMA and coeliac artery duplex ultrasound, a 50% reduction in the hepatosplenic and a 25–40% reduction in the mesenteric blood flow were demonstrated [90,91]. Simultaneous indocyanine green dye elimination measurements were

consistent with the duplex data [90]. Results from studies using gastric tonometry also support the concept of a decrease in gastrointestinal perfusion occurring with exercise. Oarsmen subjected to 30 min of maximal exercise all had a significantly reduced gastric pHi, and this was proportionally greater than the reduction in arterial pH [92]. We have produced similar results using bicycle ergometer as an exercise challenge (unpublished data) (Fig. 5).

Reduction in splanchnic blood flow occurs in proportion to relative exercise intensity. Low-intensity exercise (heart rate 90 beats/min) reduces splanchnic blood flow [93], whereas strenuous exercise can result in clinically significant gut ischaemia [94]. Increased sympathetic nervous system outflow appears to be the primary mediator of reduced blood flows to the splanchnic vasculature [95]. However, angiotensin II receptor antagonists increase blood flow throughout the gut during exercise [96], and the vasoconstrictors endothelin [48] and vasopressin probably also make important vasoconstrictive contributions [97].

Human and animal studies have shown that splanchnic blood flow is reduced less from resting levels during acute exercise after programmed endurance exercise training. The mechanisms that are involved in these adaptations produced by such training include reductions in sympathetic nervous system outflow, plasma angiotensin II and vasopressin concentrations, which result in less splanchnic and renal vasoconstriction [97].

An excellent illustration of how important adaptive splanchnic circulatory changes are during normal function is provided by studies that examine how splanchnic circulatory changes during digestion are affected by exercise. In healthy adults, exercise performed during the digestion phase does not affect the intestinal hyperaemia seen due to increased splanchnic flow. Using this knowledge has helped to provide a strategy for avoiding postprandial hypotension in elderly patients, an important clinical problem given its estimated high incidence (8% of syncopal episodes) and strong association with increased incidence of falls, syncope, new coronary events, new strokes and total mortality at long-term follow up [98]. As predicted from our knowledge of splanchnic circulation during feeding, acute exercise should ameliorate the pooling of splanchnic blood during feeding, and hence reduce postprandial hypotension. Indeed, in nursing home residents with postprandial hypotension, postprandial walking transiently increased cardiac output, but orthostatic hypotension or falls did not occur [99].

#### **Critical care**

How does normal homeostasis of gut perfusion alter in high-risk surgery or critical care diseases? Adequate splanchnic perfusion is seriously challenged during the commonly encountered critical care scenarios of circulatory shock and septic shock (like) states, but for rather different underlying reasons. Of course, clinically these two scenarios also often overlap in a dynamic manner, and therefore many ICU scenarios are likely to be considerably more complicated than the models discussed here. Unless stated otherwise, all studies quoted were conducted in animals.

#### Circulatory shock

In contrast to sepsis, circulatory shock causes splanchnic hypoperfusion with no initial change in splanchnic oxygen consumption, regardless of whether the aetiology is cardiac or acute hypovolaemia. By diverting blood supply mediated by sympathetic adrenergic stimulation [100], both the liver (which can redistribute an additional 1 l of blood to the systemic circulation under cardiovascular stress) and the gut are an efficient means of ensuring that vital organs are perfused during acute hypovolaemia [101,102], illustrating much the same general principle as that of exercise. Gastric tonometry during induced shortterm hypovolaemia in healthy volunteers demonstrated a reduced gastric pHi and this resolved with resuscitation [63]. Interestingly, this was the only significant clinical indicator of hypovolaemia, with heart rate, blood pressure and peripheral perfusion showing no change after a 20-25% blood volume venesection. Although simulated [103] and actual hypovolaemia [101] in healthy human volunteers showed that splanchnic vasoconstriction exists beyond the period of restoration of normal systemic haemodynamics after apparently adequate fluid resuscitation, the importance of the duration of such insults remains unclear. The question remains as to the cost the gut incurs as a result of sustained redistribution, or extreme hypoperfusion.

Furthermore, there is some evidence that intriguingly suggests that splanchnic hypoperfusion may actually be the result of, or at least exacerbated by, the combination of the neural response to injury plus haemorrhage, rather than hypovolaemia alone [104]. The degree of haemorrhage is increased in the presence of afferent nerve stimulation [105]. Whether afferent nerve stimulation in these animal studies simulates the pain/stress response and/or modulates central cardiovascular and baroreflexes [106] requires further investigation. It is interesting, however, to consider the obvious clinical correlate of this idea, in which neural blockade by local nerve block has been shown to reduce blood loss [107] and epidural blockade to improve lower limb graft survival [108]. However, thoracic epidural analgesia during major vascular surgery does not improve splanchnic perfusion, as monitored by gastric and sigmoid colon tonometry [109]. Laboratory studies indicate that the site and spread of epidural block are critical, with thoracic sympathetic blockade causing either no change in splanchnic perfusion [110] or increased splanchnic venodilatation, whereas lumbar blockade increases splanchnic sympathetic outflow and hence vasoconstriction, probably via the baroreceptor reflex [111].

At the local and cellular levels, the complex interplay between those factors described above in determining normal vascular tone probably plays the key role in determining whether the response to hypoperfusion is sustained. The splanchnic circulation certainly has important humorally mediated differences in response to hypoperfusion compared with the systemic circulation. Both vasopressin and angiotensin II have markedly greater effect in the mesenteric bed than elsewhere. In particular, angiotensin II is believed to play a crucial role in mediating intense splanchnic vasoconstriction. This appears to be regardless of whether the underlying aetiology is cardiogenic or haemorrhagic [112]. Splanchnic vasoconstriction is not abolished by ablation of the α-adrenergic sympathetic response [113] or mesenteric arterial denervation Nephrectomy [115], angiotensin-converting [114]. enzyme (ACE) [112,115] and specific angiotensin II receptor inhibition [116,117] prevent splanchnic vasoconstriction. Furthermore, direct infusion of angiotensin II in rats [118] and angiotensin I in humans causes splanchnic vasoconstriction, which is again reversed by ACE inhibition [119]. No clinical studies have been able to show a similar effect of ACE inhibitors on splanchnic perfusion. Splanchnic perfusion as determined by gastric tonometery was not altered by enalaprilat in adults [120] or captopril in infants [121] after cardiac surgery, although a small study of trauma patients [122] did show a benefit. As alluded to above, the mechanisms(s) that underlie such acute changes may be very different from those that are involved in chronic adaptation to low-flow states. In patients with biventricular cardiac failure, ACE inhibitors have no effect on splanchnic blood flow [123].

Vasopressin also plays an important role in producing the vasoconstriction that is seen in haemorrhage [113,124], even at relatively low levels of hypovolaemia with sympathetic blockade [125]. Antagonism of vasopressin results in higher SMA blood flow during haemorrhage [124]. Experimental models of hypovolaemia in humans, induced by head-up tilt, lower body negative pressure or epidural, show marked plasma vasopressin increases, but temporally these occur only after renin-angiotensin activation [126]. β-Blockade, which has been shown to reduce myocardial ischaemia and improve perioperative and postoperative outcome [127], elevates vasopressin levels in both euvolaemic and hypovolaemic rats [128]. Clonidine, which has also been shown to reduce myocardial ischaemia perioperatively [129], prevents sympathetic-mediated splanchnic vasoconstriction [130], including in humans [131]. Given that the splanchnic perfusion is the last to be restored after adequate fluid resuscitation, cellular changes induced by hypoperfusion that outlast the period of insult may well be the key step in perpetuating mucosal gut damage, persistent gut dysfunction and the generation of a systemic inflammatory response. The importance of reperfusion injury in the gut is reviewed elsewhere [132].

#### Sepsis/systemic inflammatory response syndrome

The hallmark of the human splanchnic circulation in sepsis/SIRS is increased total hepatosplanchnic blood flow [133–135], with higher splanchnic oxygen extraction [136] and consumption [137]. The effect on mesenteric blood flow *per se* is less clear [138]. However, oxygen consumption, delivery and extraction ratio may also depend on the duration of the sepsis state, as indicated by an endotoxin model. Oxygen consumption and delivery (but not oxygen extraction ratio) in the small intestine increased early in sepsis, only to decrease 20 h after the onset of sepsis [139]. A confounding factor in that study was the likely lack of fluid resuscitation, resulting in a haematocrit rise that may well be deleterious to the splanchnic microcirculation.

'Cytopathic hypoxia' causes this increase in hepatosplanchnic blood flow during sepsis, and many possible cellular mechanisms have been postulated [140,141]. Whether generated by initial hypoperfusion, trauma or even direct endotoxaemic damage [142], the resultant panendothelial injury that alters endothelial-derived functions generates and perpetuates an inflammatory response [143].

One problem in building an overall picture of the pathophysiology of sepsis/SIRS is the variety of methods that are employed in generating the sepsis-like response in laboratory models. These differences are probably important, particularly in interpreting the role and modulation of the splanchnic vasculature. Indeed, many endotoxin-mediated models of sepsis show decreased mesenteric perfusion, in contrast to models in which live bacterial innoculation results in the hyperdynamic response. However, in vitro, both Escherichia coli haemolysin and endotoxin models of sepsis produce abnormal capillary blood flow distribution, with decreased perfused capillaries [144], with evidence of impaired tissue oxygenation indicated by an increase in the mucosal-arterial Pco2 gap, despite the maintenance of mesenteric oxygen delivery [145]. Furthermore, relatively increased haemoglobin concentration values and oedema formation occurred, suggesting postcapillary vasoconstriction and capillary leakage. This attractive model [145] serves to illustrate the probable microvascular changes that are induced by sepsis, which are supported by current understanding of the splanchnic microvasculature [53].

Both animal and human studies show that norepinephrine, NO [146], endothelin [147] and angiotensin II [148] levels are markedly elevated in sepsis/SIRS. In addition, other vasoactive mediators such as vasoactive intestinal peptide [149], eicosanoids, platelet-activating factor and bradykinin have been implicated, but, despite successful modulation of these factors in animal models of sepsis, results in humans are disappointing [150]. In particular, NO is a key element in

generating the septic response [151,152]. However, nonspecific inhibition of NO in humans on a large scale did not improve outcome (unpublished data). NO inhibition reverses hypotension, but cardiac output is reduced and the overall effect on organ perfusion is unclear [153]. This is despite the often impressive (but also inconsistent) effects of NO inhibitors on reversing systemic hypotension and splanchnic hypoperfusion in many bacterial and endotoxin models of sepsis [154-156]. This mirrors the finding that blockade of NO synthase or gene deletion of NO synthase can exacerbate intestinal inflammation in experimental models, due to the indiscriminate inhibition of both inducible and constitutive NO [157,158]. The effect of timing of these interventions on restoring splanchnic perfusion seems important [159], given that experimental intestinal dysfunction results in an early (within 20 min) [160], severe decrease in endothelium-derived NO [161]. In contrast to the scenario of decreased NO, greater NO production is thought be responsible for decreased norepinephrine-mediated vasopressor activity during septic shock [162].

In septic patients, the greatest severity of disease has been correlated to the highest concentration of endothelin-1 [163], although the mechanism involved in the increase of endothelin-1 concentration during sepsis is largely unknown. Oldner et al [164] have shown that bosentan (a nonpeptide endothelin [ET]A and ETB receptor antagonist) restores both systemic and gut oxygen in a porcine endotoxic model of sepsis. In this model, gut oxygen consumption increased despite the profound reduction in gut oxygen delivery. Restoration of splanchnic oxygen delivery in response to bosentan treatment was not associated with an increase in oxygen consumption, suggesting that oxygen consumption was not dependent on oxygen delivery in the gut. However, that study contrasts with others conducted over longer periods of time in that, although splanchnic perfusion per se was not investigated, endothelin antagonists exacerbated endotoxic mediated hypotension [165] and increased mortality [166]. As illustrated by quantative assessments of NO and vasopressin during prolonged exposure to endotoxin or bacteria, cellular function may be profoundly altered, thereby producing a markedly different, and therefore incomparable, picture to the early response.

Given the marked increase also seen in angiotensin II, there is interesting, albeit limited, evidence that both vasopressin and angiotensin may induce endothelin release from *in vitro* endothelial cells [167]. In addition, ACE inhibitors potentiate forearm vasculature dilatation induced by infusion of acetylcholine in healthy human volunteers [168]. Although vasopressin has little effect in normal humans, its role in sepsis is complex. The effect on splanchnic perfusion of the large initial rise and subsequent rapid decline in plasma vasopressin seen in animal models is not known. Finally, impaired autonomic function

in sepsis [169], if present, has a potentially important, deleterious influence on splanchnic perfusion. The effects of inotropes and vasopressors on gastrointestinal perfusion are complex and tonometry-derived results are often not consistent with results obtained using other techniques or laboratory models. This area has been well covered in a recent review [170].

# Can improvement in gastrointestinal perfusion improve outcome?

That the majority of studies using tonometry predict outcome is certainly striking. This is all the more impressive given the inability of the commonly measured physiological variables (cardiac output and oxygen delivery excepted) to predict outcome. However, to date there is little convincing evidence that gastric tonometry-driven therapeutic intervention can alter outcome. An early study conducted in patients newly admitted to an ICU [8] demonstrated that the application of a protocol designed to increase oxygen delivery in response to an abnormal gastric pHi improved outcome in those with a normal pHi on admission, but had no effect on those who already had an abnormal pHi.

Commencement of corrective therapy in the already critically ill patient, by definition after a major insult, is considered by many to be equivalent to shutting the stable door after the horse has bolted; otherwise stated, prevention of (or rapid response to) an abnormal pHi (before or early in the course of the insult) may be much more effective than attempting to treat an established abnormal pHi. In cardiac surgical patients intraoperative oesophageal Doppler optimization of stroke volume reduced the incidence of an abnormal gastric pHi when compared with standard fluid management [6]. Whether this algorithm would improve outcome if instituted only in response to abnormal pHi is unknown. Intervention in response to an abnormal pHi that occurs intraoperatively in patients undergoing elective infrarenal aortic aneurysm repair did not improve outcome [171]. The treatment protocol in this study was somewhat different, however, and was not demonstrated to improve pHi. The hypothesis that pHiguided therapy can improve outcome was therefore not tested. One complex study that combined optimization of oxygen delivery and tonometry guidance [172] suggested that monitoring and reacting to abnormal gut perfusion produced greater benefit than that obtained by optimizing oxygen delivery alone. More recently, a large randomized controlled trial of patients recruited on ICU admission used additional fluid therapy and dobutamine in response to a low pHi, and failed to demonstrate any outcome difference between control and protocol groups [173]. Once again the treatment algorithm was ineffective at improving pHi and the therapy was instituted after the insult. An adequately powered intraoperative study with an effective treatment algorithm is awaited to answer this important clinical question definitively.

#### Conclusion

Current physiological understanding of splanchnic perfusion suggests a key role for the splanchnic circulation in the regulation of cardiovascular homeostasis. Gastrointestinal perfusion is often compromised early relative to other vascular beds in situations including critical illness, major surgery and exercise, all of which are characterized by increased demands on the circulation to maintain tissue oxygen delivery. Perhaps more importantly, this relative hypoperfusion often outlasts the period of the hypovolaemic insult or low-flow state. The relationship between gastric tonometry and gastrointestinal perfusion is complex. However, this is the only currently available clinically practical monitor that we have. The ability of pHi to predict outcome has repeatedly been demonstrated. CO<sub>o</sub> gap, now the accepted variable, has not yet been conclusively demonstrated to have the same predictive ability. Convincing data that demonstrate the ability of tonometryguided therapy to improve outcome remains elusive.

The striking association between outcome and gastric pHi provides an important clue to the pathophysiology of critical illness. Greater understanding of the complex physiological basis of gastrointestinal perfusion during both health and disease will hopefully open up further potential therapeutic avenues.

#### References

- Fine J, Frank ED, Ravin HA: The bacterial factor in traumatic shock. N Engl J Med 1959. 260:214–220.
- Schweinburg F, Fine J: Evidence for a lethal endotoxaemia as the fundamental feature of irreversibility in three types of traumatic shock. J Exp Med 1960, 112:793–800.
- Dantzker D: The gastrointestinal tract. The canary of the body? JAMA 1993, 270:1247-1248.
- Pastores SM, Katz DP, Kvetan V: Splanchnic ischemia and gut mucosal injury in sepsis and the multiple organ dysfunction syndrome. Am J Gastroenterol 1996; 91:1697–1710.
- Bennett-Guerrero E, Welsby I, Dunn TJ, Young LR, Wahl TA, Diers TL, Phillips-Bute BG, Newman MF, Mythen MG: The use of a postoperative morbidity survey to evaluate patients with prolonged hospitalization after routine, moderate-risk, elective surgery. Anesth Analg 1999; 89:514–519.
- Mythen MG, Webb AR: Perioperative plasma volume expansion reduces the incidence of gut mucosal hypoperfusion during cardiac surgery. Arch Surg 1995, 130:423–429.
- Doglio GR, Pusajo JF, Egurrola MA, Bonfigli GC, Parra C, Vetere L, Hernandez MS, Fernandez S, Palizas F, Gutierrez G: Gastric mucosal pH as a prognostic index of mortality in critically ill patients. Crit Care Med 1991, 19:1037–1041.
- Gutierrez G, Palizas F, Doglio G, Wainsztein N, Gallesio A, Pacin J, Dubin A, Schiavi E, Jorge M, Pusajo J, et al: Gastric intramucosal pH as a therapeutic index of tissue oxygenation in critically ill patients. Lancet 1992, 339:195–199.
- Fiddian-Green RG, McGough E, Pittenger G, Rothman E: Predictive value of intramural pH and other risk factors for massive bleeding from stress ulceration. Gastroenterology 1983, 85:613–620.
- Marik PE: Gastric Intramucosal pH: a better predictor of multiorgan dysfunction syndrome and death than oxygen-derived variables in patients with sepsis. Chest 1993, 104:225-229.
- Maynard N, Bihari D, Beale R, Smithies M, Baldock G, Mason R, McColl I: Assessment of splanchnic oxygenation by gastric tonometry in patients with acute circulatory failure. *JAMA* 1993, 270:1203–1210.
- Uusaro A, Ruokonen E, Takala J: Estimation of splanchnic blood flow by the Fick principle in man and problems in the use of indocyanine green. Cardiovasc Res 1995, 30:106–112.

- Landow L, Phillips DA, Heard SO, Prevost D, Vandersalm TJ, Fink MP: Gastric tonometry and venous oximetry in cardiac surgery patients. Crit Care Med 1991. 19:1226–1233.
- Ahn H, Ivarsson LE, Johansson K, Lindhagen J, Lundgren O: Assessment of gastric blood flow with laser Doppler flowmetry. Scand J Gastroenterol 1988, 23:1203–1210.
- Leung FW, Morishita T, Livingston EH, Reedy T, Guth PH: Reflectance spectrophotometry for the assessment of gastroduodenal mucosal perfusion. Am J Physiol 1987, 252:G797–G804.
- Larsen PN, Moesgaard F, Naver L, Rosenberg J, Gottrup F, Kirkegaard P, Helledie N: Gastric and colonic oxygen tension measured with a vacuum-fixed oxygen electrode. Scand J Gastroenterol 1991, 26: 409–418
- DeNobile J, Guzzetta P, Patterson K: Pulse oximetry as a means of assessing bowel viability. J Surg Res 1990, 48:21–23.
- Glenny RW, Bernard S, Brinkley M: Validation of fluorescentlabeled microspheres for measurement of regional organ perfusion. J Appl Physiol 1993, 74:2585–2597.
- Larson MV, Ahlquist DA, Karlstrom L, Sarr MG: Intraluminal measurement of enteric mucosal perfusion: relationship to superior mesenteric artery flow during basal and postprandial states in the dog. Surgery 1994, 115:118–126.
- Chapman MV, Woolf RL, Mythen MG: Temperature dependant errors using gastrointestinal tonometry: an in vivo study [abstract 580]. Br J Anaesth 1999, 82:174.
- Jakob SM, Kosonen P, Ruokonen E, Parviainen I, Takala J. The Haldane effect: an alternative explanation for increasing gastric mucosal PCO2 gradients? Br J Anaesth 1999, 83:740-746.
- Noone RB Jr, Mythen MG, Vaslef SN: Effect of alpha(alpha)-cross-linked hemoglobin and pyridoxalated hemoglobin polyoxyethylene conjugate solutions on gastrointestinal regional perfusion in hemorrhagic shock. J Trauma Injury Infect Crit Care 1998, 45:457-469
- Chapman MV, Mythen MG, Webb AR, Vincent JL: Gastrointestinal tonometry: state of the art. Intensive Care Med 2000, 26:613–622.
- Ohri SK, Bowles CT, Siddiqui A, Khaghani A, Keogh BE, Wright G, Yacoub MH, Taylor KM: The effect of cardiopulmonary bypass on gastric and colonic mucosal perfusion: a tonometric assessment. Perfusion 1994, 9:101–108.
- Guzman JA, Lacoma FJ, Kruse JA: Gastric and esophageal intramucosal PCO2 (PiCO2) during endotoxemia: assessment of raw PiCO2 and PCO2 gradients as indicators of hypoperfusion in a canine model of septic shock. Chest 1998, 113:1078-1083.
- Weil MH, Nakagawa Y, Tang W, Sato Y, Ercoli F, Finegan R, et al: Sublingual capnometry: a new noninvasive measurement for diagnosis and quantitation of severity of circulatory shock. Crit Care Med 1999, 27:1225–1229.
- Rowell LB, Detry JM, Blackmon JR, Wyss C: Importance of the splanchnic vascular bed in human blood pressure regulation. J Appl Physiol 1972. 32:213–220.
- Richardson PD, Withrington PG: Physiological regulation of the hepatic circulation. Annu Rev Physiol 1982, 44:57–69.
- McCuskey RS: Hepatic and splanchnic microvascular responses to inflammation and shock. Hepatogastroenterology 1999, 46 (Suppl 2):1464-1467.
- Bender A: The effect of increasing age on the distribution of peripheral blood flow in man. J Am Geriatr Soc 1965, 13:61-65.
- Rosenblum JD, Boyle CM, Schwartz LB: The mesenteric circulation.
  Anatomy and physiology. Surg Clin North Am 1997, 77:289–306.
- Folkow B: Regional adjustments of intestinal blood flow. Gastroenterology 1967, 52:423–432.
- Granger DN, Richardson PD, Kvietys PR, Mortillaro NA: Intestinal blood flow. Gastroenterology 1980, 78:837–863.
- Ralevic V: Splanchnic circulatory physiology. Hepatogastroenterology 1999, 46(Suppl 2):1409–1413.
- Von Kugelgen I, Starke K: Noradrenaline and adenosine triphosphate as co-transmitters of neurogenic vasoconstriction in rabbit mesenteric artery. J Physiol 1985, 367:435–455.
- Donoso MV, Brown N, Carrasco C, Cortes V, Fournier A, Huidobro-Toro JP: Stimulation of the sympathetic perimesenteric arterial nerves releases neuropeptide Y potentiating the vasomotor activity of noradrenaline: involvement of neuropeptide Y-Y1 receptors. J Neurochem 1997, 69:1048–1059.
- Han S, Yang CL, Chen X, Naes L, Cox BF, Westfall T: Direct evidence for the role of neuropeptide Y in sympathetic nerve stimulation-induced vasoconstriction. Am J Physiol 1998, 274:H290–H294.

- Kawasaki H, Takasaki K, Saito A, Goto K: Calcitonin gene-related peptide acts a novel vasodilator transmitter in mesenteric resistance vessels of the rat. Nature 1988. 335:164–167.
- Bredt DS, Hwang PM, Snyder SH: Localization of nitric oxide synthase indicating a neural role for nitric oxide. Nature 1990, 347:768-770.
- Ahlborg G, Lundberg JM: Nitric oxide-endothelin-1 interaction in humans. J Appl Physiol 1997, 82:1593–1600.
- Boulanger C, Luscher TF: Release of endothelin from the porcine aorta: inhibition by endothlium-derived nitric oxide. J Clin Invest 1990, 85:587–590.
- Lerman A, Sandok EK, Hildebrand FL Jr, Burnett JC Jr: Inhibition of endothelium-derived relaxing factor enhances endothelin-mediated vasoconstriction. Circulation 1992, 85:1894–1898.
- Haynes W, Webb DJ: Contribution of endogenous generation of endothelin-1 to basal vascular tone. Lancet 1994, 344:852–854.
- McMillen M, Sumpio BE: Endothelins: polyfunctional cytokines. J Am Coll Surg 1995, 180:621–637.
- Gardiner SM, Kemp PA, March JE, Bennett T: Effects of bosentan (Ro 47-0203), an ETA-, ETB-receptor antagonist, on regional haemodynamic responses to endothelins in conscious rats. Br J Pharmacol 1994, 112:823-830.
- Richard VHM, Clozel M, Loffler B, Thuillez C: In vivo evidence of an endothelin-induced vasopressor tone after inhibition of nitric oxide synthesis in rats. Circulation 1995, 91:771–775.
- Kubes P: Nitric oxide modulates epithlial permeability in the feline small intestine. Am J Physiol 1992, 262:G1138–G1142.
- Ahlborg G, Weitzberg É, Lundberg JM: Circulating endothelin-1 reduces splanchnic and renal blood flow and splanchnic glucose production in humans. J Appl Physiol 1995, 79:141–145.
- Burnstock G, Ralevic V: New insights into the local regulation of blood flow by perivascular nerves and endothelium. Br J Plastic Surg 1994, 47:527-543.
- Kawasaki H, Urabe M, Takasaki K: Presynaptic alpha 2-adrenoceptor modulation of 5-hydroxytryptamine and noradrenaline release from vascular adrenergic nerves. Eur J Pharmacol 1989, 164:35–43.
- Rubino A, Ralevic V, Burnstock G: The P1-purinoceptors that mediate the prejunctional inhibitory effect of adenosine on capsaicin-sensitive nonadrenergic noncholinergic neurotransmission in the rat mesenteric arterial bed are of the A1 subtype. J Pharmacol Exp Ther 1993, 267:1100-1104.
- Hamulu A, Atay Y, Yagdi T, Discigil B, Bakalim T, Buket S, Bilkay O: Effects of flow types in cardiopulmonary bypass on gastric intramucosal pH. Perfusion 1998, 13:129–135. (Published erratum appears in Perfusion 1998, 13:155.)
- Lee J-S: Biomechanics of the circulation, an integrative and therapeutic perspective. Ann Biomed Eng 2000, 28:1–13.
- Reilly PM, Bulkley GB: Vasoactive mediators and splanchnic perfusion. Crit Care Med 1993, 21(2 Suppl):S55–S68.
- Lundgren O, Svanvik J: Mucosal hemodynamics in the small intestine of the cat during reduced perfusion pressure. Acta Physiol Scand 1973, 88:551–563.
- Takala J: Determinants of splanchnic blood flow. Br J Anaesth 1996. 77:50–58.
- Kvietys PR, Granger DN: Relation between intestinal blood flow and oxygen uptake. Am J Physiol 1982, 242:G202–G208.
- Granger HJ, Norris CP: Intrinsic regulation of intestinal oxygenation in the anesthetized dog. Am J Physiol 1980, 238:H836–H843.
- Bulkley GB, Womack WA, Downey JM, Kvietys PR, Granger DN: Characterization of segmental collateral blood flow in the small intestine. Am J Physiol 1985, 249:G228–G235.
- Granger HJ, Shepherd AP Jr: Intrinsic microvascular control of tissue oxygen delivery. Microvasc Res 1973, 5:49–72.
- Bulkley GB, Kvietys PR, Parks DA, Perry MA, Granger DN: Relationship of blood flow and oxygen consumption to ischemic injury in the canine small intestine. Gastroenterology 1985, 89:852–857.
- Guzman JA, Lacoma FJ, Kruse JA: Relationship between systemic oxygen supply dependency and gastric intramucosal PCO2 during progressive hemorrhage. J Trauma 1998, 44:696-700.
- Hamilton-Davies C, Mythen MG, Salmon JB, Jacobson D, Shukla A, Webb AR: Comparison of commonly used clinical indicators of hypovolaemia with gastrointestinal tonometry. *Intensive Care Med* 1997. 23:276–281.
- Bacher A, Mayer N, Rajek AM, Haider W: Acute normovolaemic haemodilution does not aggravate gastric mucosal acidosis during cardiac surgery. Intensive Care Med 1998, 24:313–321.

- Lipsitz LA, Ryan SM, Parker JA, Freeman R, Wei JY, Goldberger AL: Hemodynamic and autonomic nervous system responses to mixed meal ingestion in healthy young and old subjects and dysautonomic patients with postprandial hypotension. Circulation 1993. 87:391–400.
- Kooner JS, Raimbach S, Watson L, Bannister R, Peart S, Mathias CJ: Relationship between splanchnic vasodilation and postprandial hypotension in patients with primary autonomic failure. J Hypertens 1989, 7(Suppl):S40-S41.
- Parker DR, Carlisle K, Cowan FJ, Corrall RJ, Read AE: Postprandial mesenteric blood flow in humans: relationship to endogenous gastrointestinal hormone secretion and energy content of food. Eur J Gastroenterol Hepatol 1995, 7:435–440.
- Maule S, Chaudhuri KR, Thomaides T, Pavitt D, McCleery J, Mathias CJ: Effects of oral alcohol on superior mesenteric artery blood flow in normal man, horizontal and tilted. Clin Sci (Colch) 1993, 84:419–425.
- Meehan AG, Kreulen DL: A capsaicin-sensitive inhibitory reflex from the colon to mesenteric arteries in the guinea-pig. J Physiol (Lond) 1992, 448:153–159.
- Geelkerken RH, Lamers CB, Delahunt TA, Hermans J, Zwijsen JH, van Bockel JH, Duodenal meal stimulation leads to coeliac artery vasoconstriction and superior mesenteric artery vasodilatation: an intra-abdominal ultrasound study. Ultrasound Med Biol 1998, 24: 1351–1356.
- Langkamp-Henken B, Kudsk KA, Proctor KG: Fasting-induced reduction of intestinal reperfusion injury. J Parent Ent Nutr 1995, 19:127–132.
- Holst JJ, Fahrenkrug J, Stadil F, Rehfeld JF: Gastrointestinal endocrinology. Scand J Gastroenterol Suppl 1996, 216:27–38.
- Mailman D: Effects of vasoactive intestinal polypeptide on intestinal absorption and blood flow. J Physiol (Lond) 1978, 279:121–132.
- Schuurkes JA, Charbon GA: Motility and hemodynamics of the canine gastrointestinal tract. Stimulation by pentagastrin, cholecystokinin and vasopressin. Arch Int Pharmacodyn Ther 1978, 236: 214–227.
- 75. Chou CC: Splanchnic and overall cardiovascular hemodynamics during eating and digestion. Fed Proc 1983, 42:1658–1661.
- Kokko JP: Countercurrent exchanger in the small intestine of man: is there evidence for its existence? Gastroenterology 1978, 74: 791–793.
- Bustamante SA, Jodal M, Nilsson NJ, Lundgren O: Evidence for a countercurrent exchanger in the intestinal villi of suckling swine. Acta Physiol Scand 1989, 137:207–213.
- Eade MN, Pybus J, Ready J: No evidence of a countercurrent multiplier in the intestinal villus of the dog. Gastroenterology 1990, 98: 3–10
- Aranow JS, Fink MP: Determinants of intestinal barrier failure in critical illness. Br J Anaesth 1996, 77:71–81.
- Lundgren O: Studies on blood flow distribution and countercurrent exchange in the small intestine. Acta Physiol Scand 1967, 303 (Suppl):1-42.
- Shepherd AP, Kiel JW: A model of countercurrent shunting of oxygen in the intestinal villus. Am J Physiol 1992, 262:H1136– H1142.
- Kiel JW, Riedel GL, Shepherd AP: Effects of hemodilution on gastric and intestinal oxygenation. Am J Physiol 1989, 256:H171– H178
- 83. Jodal M, Lundgren O: Regional distribution of red cells, plasma and blood volume in the intestinal wall of the cat. *Acta Physiol Scand* 1970, 80:533–537.
- Kolkman JJ, Groeneveld AB, van der Berg FG, Rauwerda JA, Meuwissen SG: Increased gastric PCO2 during exercise is indicative of gastric ischaemia: a tonometric study. Gut 1999, 44:163–167.
- Moore FA, Moore EE: The benefits of enteric feeding. Adv Surg 1996, 30:141-154.
- Shikora SA, Ogawa AM: Enteral nutrition and the critically ill. Postgrad Med J 1996, 72:395–402.
- Moore FA, Moore EE, Jones TN, McCroskey BL, Peterson VM: TEN versus TPN following major abdominal trauma: reduced septic morbidity. J Trauma Injury Infect Crit Care 1989, 29:916–922, discussion 922–923.
- Lawlor DK, Inculet RI, Malthaner RA: Small-bowel necrosis associated with jejunal tube feeding. Can J Surg 1998, 41:459–462.
- Caru B, Colombo E, Santoro F, Laporta A, Maslowsky F: Regional flow responses to exercise. Chest 1992, 101(Suppl 5):223S– 225S.

- Perko MJ, Nielsen HB, Skak C, Clemmesen JO, Schroeder TV, Secher NH: Mesenteric, coeliac and splanchnic blood flow in humans during exercise. J Physiol (Lond) 1998, 513:907–913.
- 91. Qamar MI, Read AE: The effects of exercise on mesenteric blood flow in man. Gut 1987, 28:583-587.
- Nielsen HB, Svendsen LB, Jensen TH, Secher NH: Exercise-induced gastric mucosal acidosis. Med Sci Sports Exerc 1995, 27:1003–1006.
- Osada T, Katsumura T, Hamaoka T, Inoue S, Esaki K, Sakamoto A, Murase N, Kajiyama J, Shimomitsu T, Iwane H: Reduced blood flow in abdominal viscera measured by Doppler ultrasound during onelegged knee extension. J Appl Physiol 1999, 86:709–719.
- Halvorsen FA, Ritland S: Gastrointestinal problems related to endurance event training. Sports Med 1992, 14:157–163.
- Kinugawa T, Ogino K, Kitamura H, Miyakoda H, Saitoh M, Hasegawa J, Kotake H, Mashiba H: Response of sympathetic nervous system activity to exercise in patients with congestive heart failure. Eur J Clin Invest 1991, 21:542-547.
- Stebbins CL, Symons JD: Role of angiotensin II in hemodynamic responses to dynamic exercise in miniswine. J Appl Physiol 1995, 78:185–190.
- McAllister RM: Adaptations in control of blood flow with training: splanchnic and renal blood flows. Med Sci Sports Exerc 1998, 30:375-381.
- Jansen RW, Lipsitz LA: Postprandial hypotension: epidemiology, pathophysiology, and clinical management. Ann Intern Med 1995, 122:286–295
- Oberman AS, Harada RK, Gagnon MM, Kiely DK, Lipsitz LA: Effects of postprandial walking exercise on meal-related hypotension in frail elderly patients [abstract 11]. Am J Cardiol 1999, 84:1130–1132.
- Chien S: Role of the sympathetic nervous system in hemorrhage. Physiol Rev 1967, 47:214–288.
- 101. Price HL, Deutsch S, Marshall BE, Stephen GW, Behar MG, Neufeld GR: Hemodynamic and metabolic effects of hemorrhage in man, with particular reference to the splanchnic circulation. Circ Res 1966, 18:469-474.
- 102. Vatner SF: Effects of hemorrhage on regional blood flow distribution in dogs and primates. *J Clin Invest* 1974, **54**:225–235.
- 103. Edouard AR, Degremont AC, Duranteau J, Pussard E, Berdeaux A, Samii K: Heterogeneous regional vascular responses to simulated transient hypovolemia in man. Intensive Care Med 1994, 20: 414–420.
- 104. Mackway-Jones K, Foex BA, Kirkman E, Little RA: Modification of the cardiovascular response to hemorrhage by somatic afferent nerve stimulation with special reference to gut and skeletal muscle blood flow. J Trauma 1999, 47:481–485.
- 105. Overman R, Wang SC: The contributory role of the afferent nervous factor in experimental shock: sublethal hemorrhage and sciatic nerve stimulation. Am J Physiol 1947, 148:289–295.
- 106. Quest JA, Gebber GL: Modulation of baroreceptor reflexes by somatic afferent nerve stimulation. Am J Physiol 1972, 222:1251– 1259.
- 107. Stevens RD, Van Gessel E, Flory N, Fournier R, Gamulin Z: Lumbar plexus block reduces pain and blood loss associated with total hip arthroplasty. Anesthesiology 2000, 93:115-121.
- 108. Christopherson R, Beattie C, Frank SM, Norris EJ, Meinert CL, Gottlieb SO, Yates H, Rock P, Parker SD, Perler BA, et al: Perioperative morbidity in patients randomized to epidural or general anesthesia for lower extremity vascular surgery. Perioperative Ischemia Randomized Anesthesia Trial Study Group. Anesthesiology 1993, 79:422-434.
- 109. Vaisanen O, Ruokonen E, Parviainen I, Bocek P, Takala J: Ranitidine or dobutamine alone or combined has no effect on gastric intramucosal-arterial PCO(2) difference after cardiac surgery. Intensive Care Med 2000, 26:45-51.
- 110. Meissner A, Weber TP, Van Aken H, Rolf N: Limited upper thoracic epidural block and splanchnic perfusion in dogs. Anesth Analg 1999, 89:1378–1381.
- 111. Hogan QH, Stekiel TA, Stadnicka A, Bosnjak ZJ, Kampine JP: Region of epidural blockade determines sympathetic and mesenteric capacitance effects in rabbits. *Anesthesiology* 1995, 83:604–610.
- 112. Toung T, Reilly PM, Fuh KC, Ferris R, Bulkley GB: Mesenteric vasoconstriction in response to hemorrhagic shock. Shock 2000, 13:267–273.
- 113. McNeill JR, Stark RD, Greenway CV: Intestinal vasoconstriction after hemorrhage: roles of vasopressin and angiotensin. Am J Physiol 1970, 219:1342–1347.

- 114. Adar R, Franklin A, Spark RF, Rosoff CB, Salzman EW: Effect of dehydration and cardiac tamponade on superior mesenteric artery flow: role of vasoactive substances. Surgery 1976, 79:534– 543
- 115. Bailey RW, Bulkley GB, Hamilton SR, Morris JB, Haglund UH: Protection of the small intestine from nonocclusive mesenteric ischemic injury due to cardiogenic shock. Am J Surg 1987, 153:108–116.
- 116. Suvannapura A, Levens NR: Local control of mesenteric blood flow by the renin-angiotensin system. Am J Physiol 1988, 255:G267– G274.
- 117. Aneman A, Svensson M, Broome M, Biber B, Petterson A, Fandriks L: Specific angiotensin II receptor blockage improves intestinal perfusion during graded hypovolemia in pigs. Crit Care Med 2000, 28: 818–823.
- 118. Bailey RW, Bulkley GB, Hamilton SR, Morris JB, Smith GW: Pathogenesis of nonocclusive ischemic colitis. Ann Surg 1986, 203: 590–599.
- 119. Gasic S, Heinz G, Kleinbloesem C, Korn A: Effects of ACE inhibition with cilazapril on splanchnic and systemic haemodynamics in man. Br J Clin Pharmacol 1989, 27(Suppl 2):225S-234S.
- 120. Parviainen I, Rantala A, Ruokonen E, Tenhunen J, Takala J: Angiotensin converting enzyme inhibition has no effect on blood pressure and splanchnic perfusion after cardiac surgery. J Crit Care 1998, 13:73–80.
- 121. Booker PD, Davis AJ, Franks R: Gut mucosal perfusion in infants undergoing cardiopulmonary bypass: effect of preoperative captopril. Br J Anaesth 1997, 79:14-18.
- 122. Kincaid EH, Miller PR, Meredith JW, Chang MC: Enalaprilat improves gut perfusion in critically injured patients. Shock 1998, 9:79-83.
- 123. Leier CV: Regional blood flow responses to vasodilators and inotropes in congestive heart failure. Am J Cardiol 1988, 62: 86E-93E.
- 124. Hock CE, Su JY, Lefer AM: Role of AVP in maintenance of circulatory homeostasis during hemorrhagic shock. Am J Physiol 1984, 246:H174-H179.
- Korner Pl, Oliver JR, Zhu JL, Gipps J, Hanneman F: Autonomic, hormonal, and local circulatory effects of hemorrhage in conscious rabbits. *Am J Physiol* 1990, 258:H229–H239.
- 126. Sander Jensen K: Heart and endocrine changes during central hypovolemia in man. Dan Med Bull 1991, 38:443-457.
- 127. Wallace A, Layug B, Tateo I, Li J, Hollenberg M, Browner W, Miller D, Mangano DT, et al: Prophylactic atenolol reduces postoperative myocardial ischemia. McSPI Research Group. Anesthesiology 1998. 88:7–17.
- 128. Peysner K, Forsling ML: Vasopressin release in response to hypovolaemia in the conscious rat and the effect of opioid and aminer-gic receptor antagonists. J Physiol Pharmacol 1991, 42:317–326.
- 129. McSPI-Europe Research Group: Perioperative sympatholysis. Beneficial effects of the alpha 2-adrenoceptor agonist mivazerol on hemodynamic stability and myocardial ischemia. Anesthesiology 1997, 86:346–363.
- 130. Yokotani K, Osumi Y: Effects of DJ-7141, a new peripherally acting alpha-2 adrenoceptor agonist, on blood pressure and gastric acid output in rats: evidence for the heterogeneity of alpha-2 adrenoceptors in different organs. *Jpn J Pharmacol* 1987, 43:415–422.
- Mathias CJ: Pharmacological manipulation of human gastrointestinal blood flow. Fundam Clin Pharmacol 1997, 11:29–34.
- 132. Biffl WL, Moore EE: Splanchnic ischaemia/reperfusion and multiple organ failure. Br J Anaesth 1996, 77:59–70.
- 133. Dahn MS, Lange P, Lobdell K, Hans B, Jacobs LA, Mitchell RA: Splanchnic and total body oxygen consumption differences in septic and injured patients. Surgery 1987, 101:69–80.
- 134. Wilmore DW, Goodwin CW, Aulick LH, Powanda MC, Mason AD Jr, Pruitt BA Jr: Effect of injury and infection on visceral metabolism and circulation. Ann Surg 1980, 192:491–504.
- 135. Gump FE, Kinney JM, Price JB Jr: Energy metabolism in surgical patients: oxygen consumption and blood flow. J Surg Res 1970, 10:613-627.
- 136. Ruokonen E, Takala J, Kari A, Saxen H, Mertsola J, Hansen EJ: Regional blood flow and oxygen transport in septic shock. Crit Care Med 1993, 21:1296-1303.
- 137. Arvidsson D, Rasmussen I, Almqvist P, Niklasson F, Haglund U: Splanchnic oxygen consumption in septic and hemorrhagic shock. Surgery 1991, 109:190-197.
- 138. Fink MP: Adequacy of gut oxygenation in endotoxemia and sepsis. Crit Care Med 1993, 21(Suppl 2):S4-S8.

- 139. Yang S, William G, Bland K, Chaudry IH, Wang P: Differential alterations in systemic and regional oxygen delivery and consumption during the early and late stages of sepsis. J Trauma 1999, 47:706.
- 140. Fink MP: Does tissue acidosis in sepsis indicate tissue hypoperfusion? *Intensive Care Med* 1996, **22**:1144–1146.
- 141. Fink M: Cytopathic hypoxia in sepsis. Acta Anaesthesiol Scand 1997, 110(Suppl):87-95.
- 142. VanderMeer TJ, Wang H, Fink MP: Endotoxemia causes ileal mucosal acidosis in the absence of mucosal hypoxia in a normodynamic porcine model of septic shock. Crit Care Med 1995, 23:1217-1226.
- 143. Wort S, Evans TW: The role of the endothelium in modulating vascular control in sepsis and related conditions. Br Med Bull 1999, 55:30–48.
- 144. Drazenovic R, Samsel RW, Wylam ME, Doerschuk CM, Schumacker PT: Regulation of perfused capillary density in canine intestinal mucosa during endotoxemia. J Appl Physiol 1992, 72:259–265.
- 145. Mayer K, Temmesfeld-Wollbruck B, Friedland A, Olschewski H, Reich M, Seeger W, Grimminger AF: Severe microcirculatory abnormalities elicited by E. coli hemolysin in the rabbit ileum mucosa. Am J Respir Crit Care Med 1999, 160:1171–1178.
- 146. Moncada S, Palmer RM, Higgs EA: Nitric oxide: physiology, pathophysiology and pharmacology. Pharmacol Rev 1991, 43:109–142.
- 147. Weitzberg E, Lundberg JM, Rudehill A: Elevated plasma levels of endothelin in patients with sepsis syndrome. Circ Shock 1991, 33: 222–227.
- 148. Aneman A, Bengtsson J, Snygg J, Holm M, Pettersson A, Fandriks L: Differentiation of the peptidergic vasoregulatory response to standardized splanchnic hypoperfusion by acute hypovolaemia or sepsis in anaesthetized pigs. Acta Physiol Scand 1999, 166:293– 300.
- 149. Broner C, O'Doriso MS, Rosenberg RB, O'Doriso TM: Cyclic nucleotides and vasoactive intestinal peptide production in a rabbit model of Escherichia coli septicemia. Am J Med Sci 1995, 309:267–277.
- 150. Fink MP: Therapeutic options directed against platelet activating factor, eicosanoids and bradykinin in sepsis. *J Antimicrob Chemother* 1998, **41(Suppl A)**:81–94.
- 151. Wort SJ, Evans TW: The role of the endothelium in modulating vascular control in sepsis and related conditions. *Br Med Bull* 1999, **55**:30–48.
- 152. Symeonides S, Balk RA: Nitric oxide in the pathogenesis of sepsis. Infect Dis Clin North Am 1999, 13:449–463.
- 153. Petros A, Lamb G, Leone A, Moncada S, Bennett D, Vallance P: Effects of a nitric oxide synthase inhibitor in humans with septic shock. Cardiovasc Res 1994, 28:34–39.
- 154. Boughton-Smith NK HI, Deakin AM, Whittle BJ, Moncada S: Protective effect of S-nitroso-N-acetyl-pencillamine in endotoxin-induced acute intestinal damage in the rat. Eur J Pharmacol 1990, 191:485–488.
- 155. Allman K, Stoddart AP, Kennedy MM, Young JD: L-Arginine augments nitric oxide production and mesenteric blood flow in ovine endotoxemia. Am J Physiol 1996, 271:H1296-H1301.
- 156. Laszlo F, Whittle BJR, Moncada S: Attenuation by nitrosothiol NO donors of acute intestinal microvascular dysfunction in the rat. Br J Pharmacol 1995, 115:498–502.
- 157. Schumacker PT, Kazaglis J, Connolly HV, Samsel RW, O'Connor MF, Umans JG: Systemic and gut O2 extraction during endotoxemia: role of nitric oxide synthesis. Am J Respir Crit Care Med 1995, 151:107–115.
- 158. Spain D, Wilson MA, Bar-Natan MF, Garrison RN: Nitric oxide synthase inhibition aggravates intestinal microvascular vasoconstriction and hypoperfusion of bacteremia. J Trauma 1994, 36: 720-725
- 159. Laszlo F, Whittle BJR, Moncada S: Time-dependent enhancement or inhibition of endotoxin-induced vascular injury in rat intestine by nitric oxide synthase inhibitors. Br J Pharmacol 1994, 111: 1309-1315.
- 160. Liu S, Barnes, PJ, Evans TW: Time course and cellular localization of lipopolysaccharide induced inducible nitric oxide synthase messenger RNA expression in the rat in vivo. Crit Care Med 1997, 25:512–518.
- 161. Lefer AM, Lefer DJ: Nitric oxide. II. Nitric oxide protects in intestinal inflammation. *Am J Physiol* 1999. **276**:G572–G575.
- 162. Tsuneyoshi I, Kanmura Y, Yoshimura N: Nitric oxide as a mediator of reduced arterial responsiveness in septic patients. Crit Care Med 1996, 24:1083–1086.

- 163. Pittet JF, Morel DR, Hemsen A, Gunning K, Lacroix JS, Suter PM, Lundberg JM: Elevated plasma endothelin-1 concentrations are associated with the severity of illness in patients with sepsis. *Ann Surg* 1991, 213:261–264.
- 164. Oldner A, Wanecek M, Weitzberg E, Rundgren M, Alving K, Ullman J, Rudehill A: Angiotensin II receptor antagonism increases gut oxygen delivery but fails to improve intestinal mucosal acidosis in porcine endotoxin shock. Shock 1999, 11:127–135.
- 165. Gardiner SM, Kemp PA, March JE, Bennett T: Enhancement of the hypotensive and vasodilator effects of endotoxaemia in conscious rats by the endothelin antagonist, SB 209670. Br J Pharmacol 1995, 116:1718–1719.
- 166. Ruetten H, Thiemermann C, Vane JR: Effects of the endothelin receptor antagonist, SB 209670, on circulatory failure and organ injury in endotoxic shock in the anaesthetized rat. Br J Pharmacol 1996, 118:198-204.
- 167. Yoshida H, Nakamura M: Inhibition by angiotensin converting enzyme inhibitors of endothelin secretion from cultured human endothelial cells. *Life Sci* 1992, **50**:L195–L200.
- 168. Nakamura M, Funakoshi T, Yoshida H, Arakawa N, Suzuki T, Hiramori K: Endothelium-dependent vasodilation is augmented by angiotensin converting enzyme inhibitors in healthy volunteers. J Cardiovasc Pharmacol 1992, 20:949–954.
- 169. Garrard CS, Kontoyannis DA, Piepoli M: Spectral analysis of heart rate variability in the sepsis syndrome. Clin Auton Res 1993, 3:5–13.
- Silva E, DeBacker D, Creteur J, Vincent JL: Effects of vasoactive drugs on gastric intramucosal pH. Crit Care Med 1998, 26:1749–1758.
- 171. Pargger H, Hampl KF, Christen P, Staender S, Scheidegger D: Gastric intramucosal pH-guided therapy of infrarenal abdominal aneurysms: is it beneficial? *Intensive Care Med* 1998, 24:769-776.
- 172. Ivatury RR, Simon RJ, Islam S, Fueg A, Rohman M, Stahl WM: A prospective randomized study of end points of resuscitation after major trauma: global oxygen transport indices versus organ-specific gastric mucosal pH. J Am Coll Surg 1996, 183:145–154.
- 173. Gomersall CD, Joynt GM, Freebairn RC, Hung V, Buckley TA, Oh TE: Resuscitation of critically ill patients based on the results of gastric tonometry: a prospective, randomized, controlled trial. Crit Care Med 2000, 28:607–614.

Authors' affiliation: Centre for Anaesthesia, University College London, London, UK

Correspondence: Michael Grocott, Centre for Anaesthesia, University College London, Room 103, The Crosspiece, Middlesex Hospital, Mortimer Street, London, W1N 8AA. Tel: +44 20 7380 9477; fax: +44 20 7580 6423; e-mail: mikegrocott@hotmail.com